Freeform Search

	US Pre-Grant Publication Full-Text Database						
	US Patents Full-Text Database						
	US OCR Full-Text Database						
Database:	EPO Abstracts Database						
	JPO Abstracts Database						
	Derwent World Patents Index						
	IBM Technical Disclosure Bulletins						
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	L7 and (hybridiz\$5 near5 probe\$1 near5						
Term:	(plurality or multiple))						
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Display:	Documents in Display Format: - Starting with Number 1	٦					
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DATE: Wednesday, January 26, 2005 Printable Copy Create Case

Set Name Query		Hit Count Set Name	
side by side			result set
DB = U	JSPT,EPAB,JPAB,DWPI; PLUR=YES; OP=ADJ		
<u>L8</u>	L7 and (hybridiz\$5 near5 probe\$1 near5 (plurality or multiple))	28	<u>L8</u>
<u>L7</u>	11 and amplifi\$2 and hybridiz\$5 and detect\$3	319	L7
<u>L6</u>	11 and amplifier\$1 and hybridizer\$1 and detector\$1	0	L6
<u>L5</u>	L4 and bacter\$3	2	L5
<u>L4</u>	12 and intestin\$2	3	L4
<u>L3</u>	L2 and intestin\$2 and bacter\$3 and flora	0	L3
<u>L2</u>	L1 and (hybridiz\$5 near5 probe\$1 near5 (plurality or multiple))	28	L2
L1	(apparatus\$1 or device\$1) near5 PCR	757	L1

END OF SEARCH HISTORY

Freeform Search

Generate:	O Hit List O Hit Count O Side by Side O	Image	
Display:	Documents in Display Format: -	Starting with Number 11	1
Term:	113 and 16S ribosomal RNA		
Database:	US Pre-Grant Publication Full-Text Database US Patents Full-Text Database US OCR Full-Text Database EPO Abstracts Database JPO Abstracts Database Derwent World Patents Index IBM Technical Disclosure Bulletins		

Search History

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DATE: Wednesday, January 26, 2005 Printable Copy Create Case

Name side by side	Query	Hit Count	Set Name result set	
DB=USPT,EPAB,JPAB,DWPI; PLUR=YES; OP=ADJ				
<u>L14</u>	113 and 16S ribosomal RNA	3	L14	
L13	L12 and (subject\$1 or patient\$1)	107	L13	
L12	L11 and (probe\$1 near5 immobiliz\$5)	112	L12	
<u>L11</u>	19 and (apparatus or device)	279	L11	
L10	L9 and 16SrRNA	0	L10	
<u>L9</u>	12 and ((plurality or multiple) near5 probe\$1)	468	L9	
<u>L8</u>	12 AND 16srRNA	1	L8	
<u>L7</u>	intestin\$3 bacterial flora same PCR same probe\$1 same hybridiz\$5	0	<u>L7</u>	
L6	intestinal bacterial flora near5 PCR	0	L6	
<u>L5</u>	L4 and (multiple or plurality)	3	L5	
L4	L3 and (hybridiz\$5 near5 probe\$1)	3	L4	
L3	(fecal or fecees or intestin\$3) near5 PCR near5 (flora or bacter\$3)	4	L3	
L8 L7 L6 L5 L4 L3	(fecal or feces or intestin\$3) and PCR and (flora or bacter\$3) and (hybridiz\$5 near5 probe\$1)	4906	<u>L2</u>	

L1 (fecal or fees or intestin\$3) and PCR and (flora or bacter\$3) and probe\$1 6195 L1

END OF SEARCH HISTORY

s intestin2 bacter###(10a)PCR(10a)probe# 2 IS NOT A RECOGNIZED COMMAND The previous command name entered was not recognized by the system. For a list of commands available to you in the current file, enter "HELP COMMANDS" at an arrow prompt (=>). s intestin## bacter###(10a)PCR(10a)probe# L14 INTESTIN## BACTER###(10A) PCR(10A) PROBE# => s l1 and (plurality or multiple) 1 L1 AND (PLURALITY OR MULTIPLE) => s 12 and flora 1 L2 AND FLORA => d 13 bib ab kwic ANSWER 1 OF 1 CAPLUS COPYRIGHT 2005 ACS on STN L3 AN 2002:853202 CAPLUS DN 138:215878 ΤI PCR-ELISA. I: Application to simultaneous analysis of mixed bacterial samples composed of intestinal species ΑU Laitinen, Reija; Malinen, Erja; Palva, Airi CS Faculty of Veterinary Medicine, Department of Basic Veterinary Sciences, Section of Microbiology, Helsinki University, Finland SO Systematic and Applied Microbiology (2002), 25(2), 241-248 CODEN: SAMIDF; ISSN: 0723-2020 PΒ Urban & Fischer Verlag GmbH & Co. KG DTJournal LA English AB Sixteen oligonucleotide identification probes, designed in this study or adapted from literature, were tested in PCR-ELISA assays for their ability to simultaneously detect under standardized conditions selected intestinal bacteria, lactobacilli and bifidobacteria. The level of specificity obtained with most of the probes fulfilled the set criteria. The lack of efficiency of PCR performed with the primers, proposed to be specific for the entire eubacteria domain, and compromises made in hybridization conditions due to simultaneous usage of multiple probes reduced the sensitivity of the PCR-ELISA test. The method was, however, found to be suitable for detecting predominant members of the intestinal Applicability of the PCR-ELISA test could be further widened using primers with a more restricted specificity in the PCR step, as was demonstrated for the detection of Bifidobacterium with genus-specific primers. Advantages of the PCR-ELISA method include convenient performance and the possibility to test rapidly large amts. of samples with a multitude of probes. RE.CNT 37 THERE ARE 37 CITED REFERENCES AVAILABLE FOR THIS RECORD ALL CITATIONS AVAILABLE IN THE RE FORMAT AB Sixteen oligonucleotide identification probes, designed in this study or adapted from literature, were tested in PCR-ELISA assays for their ability to simultaneously detect under standardized conditions selected intestinal bacteria, lactobacilli and bifidobacteria. The level of specificity obtained with most of the probes fulfilled the set criteria. The lack of efficiency of PCR performed with the primers, proposed to be specific for the entire eubacteria domain, and compromises made in hybridization conditions due to simultaneous usage of multiple probes reduced the sensitivity of the PCR-ELISA test. The method was, however, found to be suitable for detecting predominant members of the intestinal Applicability of the PCR-ELISA test could be further widened using primers with a more restricted specificity in the PCR step, as was demonstrated for the detection of Bifidobacterium with genus-specific primers. Advantages of the PCR-ELISA method include convenient performance and the possibility to test rapidly large amts. of

samples with a multitude of probes.

IT Bifidobacterium
 Intestinal bacteria
Lactobacillus
 (PCR-ELISA, using 16S or 23S rDNA-specific primers/
 probes, for simultaneous detection of intestinal bacteria, lactobacilli and bifidobacteria)